Iontophoresis—derived from the Greek ιόντος, meaning ion, and φόρεσις, meaning to move across—is a noninvasive technique in which a weak electrical current is used to transport molecules into tissue (Figure 1). Generally, a drug is applied through an electrode that carries the same charge, while the ground electrode, which carries the opposite charge, is placed elsewhere on the body to complete the circuit (Figure 2).

In certain specialities, such as gynecology and dermatology, iontophoresis has long been in use to obtain passive molecule penetration into tissues without the use of a needle. In ophthalmology, iontophoresis was first used in 1908 as a therapy for ulcers, keratitis, and episcleritis. Iontophoretic penetration of several drugs, including antibiotics, antivirals, corticosteroids, and antiphlogistics, has been established in several studies that also described dose- and time-dependency of drug penetration into the eye.

Riboflavin, being negatively charged at physiologic pH and highly soluble in water due to its small molecular weight, is an excellent candidate for iontophoresis. The idea of using iontophoresis to assist in epithelium-on (epi-on) CXL was conceived to minimize potential complications created by corneal epithelial debridement, keeping the surface intact. The possibility of side effects induced by the epithelium-off (epi-off) technique, including pain, infection, temporary vision loss, and sterile infiltrates, prompted researchers to try several approaches to deliver the same amount of riboflavin into the stroma as is achieved with epi-off; unfortunately, these approaches could not guarantee the same efficacy for stabilizing keratoconus or other ectatic diseases.

THE PROCEDURE

CXL assisted by iontophoresis is performed under topical anesthesia. A power supply, a connection cable, and a return electrode applied to the patient’s forehead are activated by a corneal applicator (Iontofor CXL; Sooft Italia) that contains negative electrodes and adheres to the cornea with a weak suction (at least 1 mL) vacuum system.

After verifying that the applicator is secured to the cornea, the operator covers the steel grid (Figure 3) with a specially formulated 0.1% riboflavin solution (Ricrolin+; Sooft Italia). The solution is hypo-osmolar, dextran-free, and enriched with ethylenediaminetetraacetic acid (ETDA) and tromethamine, which is applied for 5 minutes. During this phase, ion transport is performed by continuous erosion.

Advantages of this approach include decreased procedure time and improved riboflavin penetration.

BY LUCA GUALDI, MD; FEDERICA GUALDI, MD; VERONICA CAPPY, MD; AND MASSIMO GUALDI, MD
which allows quicker passage through the intact epithelium. The power generator is then turned on, and 1 mA power is selected to start the procedure, which takes only 5 minutes to complete.

The intensity that we found to be most effective, safe, and nontoxic was 1 mA/cm². Studies have reported side effects in vivo only at much higher voltage.

The operator must ensure that the steel grid remains covered with riboflavin solution for the duration of the procedure, thus maintaining a regular flow of electrical current. After the iontophoresis procedure, the suction is released and the corneal applicator is removed from the cornea. UV-A irradiation is then performed for 9 minutes (Figure 4).

During UV-A irradiation, balanced salt solution is applied to ensure that the corneal epithelium remains hydrated. Postoperative medications include topical antibiotics, corticosteroids, and lubricants. To reduce possible patient discomfort, a therapeutic contact lens is placed after irradiation and then removed on postoperative day 1.

One of the main advantages of iontophoresis-assisted CXL is its time efficiency: The procedure requires only 5 minutes of iontophoresis plus 9 minutes of UV-A irradiation. (However, the irradiation time following iontophoresis theoretically depends on the OXL platform used.) Early studies of hematoxylin-eosin staining and immunohistochemical analysis showed that accelerating the power from 3 to 10 mW/cm² allowed surgeons to save time and achieve better results with riboflavin penetration.

**CLINICAL EXPERIENCE**

In our experience with iontophoretic CXL, we used a commercially available UV-A optical system (CBM X-Linker Vega 10 mW; CSO). We administered one drop of pilocarpine 2% preoperatively to protect the lens and the macula from the UV-A irradiation and two or more drops of oxybuprocaine HCl 0.4% to prevent pain during the procedure. The action of the preservative included in this particular anesthetic (benzalkonium chloride) can further facilitate drug penetration into the anterior stroma because it temporarily separates the bonds between the epithelial tight junctions. This is the basis of transepithelial OXL, also known as epi-on OXL.

The evolution of this technique also includes the use of enhancers such as sodium EDTA (carboxylic acid) and tromethamine (amino-alcohol), which better facilitate riboflavin penetration.

---

**AT A GLANCE**

- Iontophoresis, a noninvasive technique in which a weak electrical current is used to transport molecules into tissue, was first used in ophthalmology in 1908 as a therapy for ulcers, keratitis, and episcleritis.

- Riboflavin, being negatively charged at physiologic pH and highly soluble in water due to its small molecular weight, is an excellent candidate for iontophoresis.

- Iontophoretic OXL has been shown to improve corneal biomechanical properties, partially halt ectatic disease progression, limit postoperative pain, minimize the risk of infection, and reduce treatment time.
The concentration of riboflavin in the stromal tissue that is needed to achieve a full crosslinking effect in keratoconus patients remains unknown. Iontophoresis is a possible way to deliver more riboflavin into the corneal tissue with the epi-on approach and to reduce the administration time.

The advantages of epi-on CXL include the absence of side effects due to epithelial debridement and resulting quick visual recovery. The procedure is completely painless, and only low doses of postoperative eye drops are required, which is extremely important for steroid responders. Few materials and drugs are consumed, and less time is needed for multiple postoperative exams. Additionally, epi-on CXL can be performed bilaterally, and it is does not necessarily have to be done in the OR. Because the technique is less invasive, it is also more easily performed in thinner corneas or in uncooperative patients, such as those affected by Down syndrome or children.

Unfortunately, the epi-on technique greatly reduces riboflavin penetration compared with the standard epi-off approach. Consequently, its efficacy has been limited. The long-term efficacy and safety of the epi-off approach have been widely demonstrated. However, patient discomfort, slow recovery, and risk of complications have also been reported. Epithelial debridement carries the inherent risks of corneal infection, subepithelial haze, sterile corneal infiltrates, corneal scarring, endothelial damage, and herpetic activation.

The aim of iontophoresis-assisted CXL is to overcome the poor efficacy of epi-on CXL while maintaining similar efficacy to the standard protocol. The early results are encouraging. In eyes treated with iontophoresis-assisted CXL, riboflavin has been seen in the posterior stroma using confocal and high-performance liquid chromatography. Additionally, the demarcation line was found at a much greater depth than with standard epi-on CXL and at a depth nearly similar to that with epi-off CXL.

Studies in human donor eyes showed an increased Young modulus of the cornea after iontophoresis-assisted CXL and a stress-strain curve highly superior to that seen with standard transepithelial (epi-on) CXL, although the biomechanical effect remained slightly inferior to that with epi-off CXL. This might be explained by a lower apoptotic effect due to reduced riboflavin supply during UV-A irradiation after the iontophoresis device is removed. Further, the amount of riboflavin absorbed through iontophoresis is greater than with the conventional epi-on protocol but lower than with the standard epi-off protocol.

The concentration of riboflavin in the stromal tissue that is needed to achieve a full crosslinking effect in keratoconus patients remains unknown. Iontophoresis is a possible way to deliver more riboflavin into the corneal tissue with the epi-on approach and to reduce the administration time.

Indications for iontophoresis-assisted CXL are nearly the same as those for standard epi-on CXL. They should include clinical and measureable progression of ectatic disease, a thinnest corneal point greater than 350 µm, and a clear cornea on biomicroscopic and confocal examination with no Vogt striae or reticular microstriae. Exclusion criteria for iontophoresis-assisted CXL include history of herpetic keratitis, current pregnancy or breastfeeding, and presence of corneal opacities.

RESULTS

We have treated eyes affected by forme fruste and mild keratoconus, eyes with pellucid marginal degeneration, and eyes with ectasia after LASIK, RK, and intrastromal corneal ring segment (ICRS) implantation (same day or 3 months after insertion). We have seen no significant side effects in any group. Four eyes with ICRSs and one eye with previous RK experienced discomfort, and punctate keratitis has been observed just after the procedure but completely resolved within 7 days. After one procedure, mild corneal edema was seen on biomicroscopy, but this completely disappeared within 2 days with application of topical steroids. In most cases, microepithelial disruption was observed, which we assumed was due to the use of enhancers (BAK, ETDA, and tromethamine).
These findings were quantified by anterior segment OCT and Scheimpflug densitometry, which confirmed complete regression within the first week in all cases. Clinically relevant endothelial cell loss was not observed. Mean endothelial cell density was 2,478 ± 318 cells/mm² preoperatively and 2,470 ± 310 cells/mm² postoperatively. No significant change in pachymetry was observed, with average thinnest point of 451 µm preoperatively and 448 µm postoperatively. Patients with corneal ectasia were treated to stabilize their disease, and no intraoperative side effects on 9-mm LASIK flaps were seen during the release of the 8-mm diameter iontophoresis suction ring.

Riboflavin penetration and fluorescence after iontophoresis is clearly visible just before and during UV-A irradiation (Figures 4 and 5). After iontophoresis-assisted CXL, in most cases, we noticed a demarcation line as a hyperreflective and nonhomogeneous band that was slightly more superficial and less intense than those obtained after epi-off CXL. The demarcation line is normally located within peaks with an average of 220 and 320 µm depth, and it appears to be more of a gradient than a defined line (Figure 6).

We evaluated 18 eyes that completed follow-up of 2 years; we subjectively measured visual acuity and objectively analyzed topography, tomography, and aberrometry. The following mean differences from baseline were observed: UCVA: -0.08 (±0.10 standard deviation [SD]), BCVA: -0.10 (±0.10 SD); average keratometry (K): -0.28 (±1.24 SD); maximum K: -0.52 (±1.68 SD); coma aberration: -0.09 µm (±0.78 SD); and total aberrations: -0.13 (±0.66 SD). These data document not only a halting of keratoconus progression but also a slight tendency to improvement in some cases, as has previously been reported.64,66

Iontophoresis-assisted CXL does not preclude eventual epi-off CXL enhancement in the event of keratoconus progression; therefore, if a cornea is suitable for other CXL procedures after iontophoresis-assisted CXL, it can be retreated using any other approach.

DISCUSSION

The aims of transepithelial CXL assisted by iontophoresis are to minimize the risk of infection, allow faster visual recovery, and decrease treatment time, all without reducing the efficacy of the procedure. We observed that this technique induces acceptable penetration of riboflavin in all corneal layers, which is the basis for efficient crosslinking, even with an intact epithelium. The effective presence of riboflavin together with UV-A produced, as observed by stress-strain measurements, a significant increase of corneal stiffness in the iontophoresis-assisted CXL group compared with controls, thereby partially confirming previous reports.69

A possible explanation of the reduced effect of iontophoresis-assisted CXL is that an intact epithelium soaked with riboflavin may partially arrest UV-A light. However, investigators have shown that the corneal epithelium limits riboflavin penetration but not UV-A light transmittance and that epithelial cells are not enriched with riboflavin.71 For that reason, only a small amount of the UV-A light, approximately 15% to 20%, should be absorbed by the epithelium.69,72

Iontophoresis studies were conducted using the same energy dose as in the standard protocol (5.4 J/cm²). Additional research is needed to determine if more irradiance power is required to compensate for the 20% UV-A light loss in the epithelium and thereby achieve the biomechanical effect of epi-off. Also still to be determined is whether the lower stiffening effect may nonetheless be enough to stabilize an ectatic cornea.64

CONCLUSION

Today, the epi-off technique is still considered the gold standard CXL procedure. Epi-on CXL, assisted by iontophoresis or not, appears to be not only a valid option but also the only way today (with contact lens-assisted CXL73) to treat corneas thinner than 400 µm.

Although longer follow-up and further studies are needed to evaluate its safety and efficacy, CXL assisted by iontophoresis is a potentially valid alternative to standard CXL. In our experience, this approach improves corneal biomechanical properties, partially halts ectatic disease progression, limits postoperative pain, minimizes the risk of infection, and reduces treatment time.


Luca Gualdi, MD  Anterior segment surgeon and Director, Studio Oculistico Gualdi, luca@gualdi.it

Luca Gualdi, MD  Anterior segment surgeon, Studio Oculistico Gualdi, luca@gualdi.it

Massimo Guidali, MD  Anterior segment surgeon and Director, Studio Oculistico Gualdi, massimo@gualdi.it

Christiane Unterhuber, MD  Clinical Eye Research Unit, University of Oxford, Oxford, United Kingdom.

Antonio Rubio, MD  Professor and Head of Ophthalmology, Select Eye Center, Valencia, Spain.

Alberto-Antonio Díaz-Cruz, MD  Associate Professor, Department of Ophthalmology, University of Chile, Santiago, Chile.

Marcus W. mimicus, MD  Associate Professor of Ophthalmology, Division of Ophthalmology, Stanford University, Stanford, California, USA.

Veronica Cappello, MD

General ophthalmologist, Studio Oculistico Gualdi, Rome

veronica.cappello@libero.it

Financial disclosure: None

Fedeaerica Guidali, MD

General ophthalmologist, Studio Oculistico Gualdi, Rome

federicaguidali@gmail.com

Financial disclosure: None

Luca Gualdi, MD

Anterior segment surgeon, Studio Oculistico Gualdi, luca@gualdi.it

Financial disclosure: None

Massimo Guidali, MD

Anterior segment surgeon and Director, Studio Oculistico Gualdi, massimo@gualdi.it

Financial disclosure: None

Cataract & Refractive Surgery Today Europe | October 2015